



07/20/00

THE COMMISSIONER OF PATENTS AND TRADEMARKS
WASHINGTON, D.C. 20231

Case Docket No.: 20092

JC598 U.S. PTO
09/620227
07/20/00

Sir:

Transmitted herewith for filing is the patent application of

Inventor: MOSHE FLUGELMANFor : ARTIFICIAL VASCULAR GRAFTS, AND METHODS OF PRODUCING AND USING SAME

Enclosed are:

- ☒ 6 sheets of informal drawing(s).
- ☐ An assignment of the invention to CCC
- ☐ A certified copy of a _____ application.
- ☐ An associate power of attorney.
- ☒ A verified statement to establish small entity status under 37 CFR 1.9 and 37 CFR 1.27.
- ☐ Other - _____

The filing fee has been calculated as shown below:

	(Col.1) NO. FILED	(Col.2) NO. EXTRA
FOR:		
BASIC FEE		
TOTAL CLAIMS	55 - 20 =	35
INDEP CLAIMS	9 - 3 =	6
... Record of Assignment		40

* If the difference in Col.1 is less
than zero, enter "0" in Col.2

SMALL ENTITY	
RATE	FEE
	\$ 345
35 x9 =	\$ 315
6 x39	\$ 234
	\$ 0
TOTAL	\$ 894

OTHER THAN A SMALL ENTITY	
RATE	FEE
	\$ 690
x18 =	\$
x78	\$
	\$
TOTAL	\$

- ☒ Please charge my Deposit Account No. 50-1407 in the amount of \$ 894. A duplicate copy of this sheet is enclosed.
- ☐ A check in the amount of \$ _____ to cover the filing fee is enclosed.
- ☒ The Commissioner is hereby authorized to charge payment of the following fees associated with this communication or credit any overpayment to Deposit Account No. 50-1407. A duplicate copy of this sheet is enclosed.
- ☒ Any additional filing fees required under 37 CFR 1.16.
- ☒ Any patent application processing fees under 37 CFR 1.17.
- ☒ The Commissioner is hereby authorized to charge payment of the following fees during the pendency of this application or credit any overpayment to Deposit Account No. 50-1407. A duplicate copy of this sheet is enclosed.
- ☒ Any patent application processing fees under 37 CFR 1.17.
- ☐ The issue fee set in 37 CFR 1.18 at or before mailing of the Notice of allowance, pursuant to 37 CFR 1.311(b).
- ☒ Any filing fees under 37 CFR 1.16 for presentation of extra claims.

Respectfully,
Sol Sheinbein
Sol Sheinbein
Reg. No. 25,457

Sol Sheinbein
G. E. Ehrlich (1995) Ltd.
C/O Anthony Castorina
2001 Jefferson Davis Highway
Suite 207
Arlington, Virginia 22202

IN THE UNITED STATES PATENT AND TRADEMARK OFFICEIn RE Application of: MOSHE FLUGELMAN

Filed Concurrently Herewith

For: ARTIFICIAL VASCULAR GRAFTS, AND METHODS OF PRODUCING AND USING SAMEVERIFIED STATEMENT UNDER 37 CFR 1.27
CLAIMING STATUS AS A SMALL ENTITY

To The Commissioner of Patents and Trademarks:

As a below named inventor, I hereby declare that:

I qualify as an independent inventor as defined in 37 CFR 1.9(c) for purposes of paying reduced fees under 35 USC § 41(a) and § 41(b) to the Patent and Trademark Office with regard to the above-entitled invention described in the specification filed herewith.

I have not assigned, granted, conveyed or licensed and am under no obligation under contract or law to assign, grant, convey or license, any rights in the invention to any party who could not qualify as a small entity under 37 CFR 1.9(f), namely any person who could not be classified as an independent inventor under 37 CFR 1.9(c) if that person had made the invention, or any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

Each party, if any, who could qualify as a small entity under 37 CFR 1.9(f) and to whom I have assigned, granted, conveyed or licensed or am under an obligation under contract or law to assign, grant, convey or license, any rights in the invention is listed below:

Full Name (Party 1) : NONE

Address : _____

Status : ☒ Individual☐ Small Business
Concern☐ Nonprofit
Organization

Full Name (Party 2) : _____

Address : _____

Status : ☒ Individual☐ Small Business
Concern☐ Nonprofit
Organization

I acknowledge the duty under 37 CFR 1.28(b) to file, in this application, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the issue fee due after the date on which status as a small entity is no longer appropriate.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application and any patent issuing thereon.


MOSHE FLUGELMANDate: 16 July, 2000

APPLICATION FOR PATENT

5 Inventor: Moshe Flugelman

Title: ARTIFICIAL VASCULAR GRAFTS, AND METHODS OF
10 PRODUCING AND USING SAME

FIELD AND BACKGROUND OF THE INVENTION

15 The present invention relates to artificial vascular grafts. More particularly, the present invention relates to genetically transformed cells and methods of generating same, which cells are useful in the generation of improved artificial vascular grafts. The present invention further relates to methods of using the artificial vascular grafts for therapy of vascular
20 diseases.

Vascular diseases:

Vascular diseases are a major cause of morbidity and mortality in the developed world and thus affects a large part of the population.

Bypass surgery utilizing venous, arterial or synthetic conduit grafts is

5 often used to alleviate the clinical sequelae of these diseases by allowing or restoring blood flow around a blocked or damaged blood vessel. It is estimated that over 1 million such procedures are performed in developed countries annually and that 30-50% of these interventions fail within 5 to 7 years (1-4).

Arterial grafts:

10 Arterial grafts have a higher durability rate in the long term when compared to venous and synthetic grafts (5). Yet, arterial grafts can be used only in limited clinical circumstances such as, for example, aorto-coronary bypass surgery. There is an ever-increasing need for synthetic

grafts with a biological-compatible surface of high durability over extended periods of time. The demand for small caliber grafts is especially high in lesions below the femoral artery and in patients who have limited native vessels such as with varicose veins or previous bypass surgery

5 (2,3). Use of polytetrafluoroethylene (PTFE) and other materials in synthetic grafts has gained popularity due to the biocompatibility of the material itself and to its durability. Modification of the synthetic material was described in many animal studies and in a few human studies (5-10).

To improve function and durability of synthetic grafts, autologous
10 endothelial cells have been used to seed grafts before implantation.

Endothelial cells provide the best bio-compatible surface, and may provide long term protection from thrombosis due to their thrombolytic capacity (11,12). In addition, endothelial cell coverage can also prevent neointimal proliferation and inflammatory reaction in the graft (13). However, to date,

randomized large scale studies have not proven that endothelial cell seeding can improve graft durability (8-10).

Endothelial cells used for seeding grafts before implantation are typically harvested from short venous segments or from adipose tissue.

5 Other sources may be cells from transgenic animals having human characteristics or autologous endothelial cells from the individual patient.

Another possible source for endothelial cells is progenitor cells isolated from the peripheral blood or bone marrow.

New strategies have been attempted at the stage of artificial graft
10 design and preparation in order to improve efficiency of artificial vascular grafts utilized in treating human vascular diseases. Complete endothelialization of artificial vascular graft luminal surface is still extremely delayed and has a major impact on graft patency. Therefor, a major task is to improve both graft coverage and cell adherence to the

graft matrix. Previous attempts in which endothelial cells were seeded onto synthetic grafts, resulted in incomplete endothelialization as well as cell detachment upon exposure to blood flow conditions.

One of the most recent techniques of improving graft endothelialization is to coat it with genetically transformed cells having better proliferating capacity.

To enhance differentiation of progenitor bone marrow cells and provide a short and reliable method for generating endothelial cells useable in artificial graft preparation, such cells are typically transformed with a vascular endothelial growth factor (VEGF) expression vector which when expressed in the cell, can enhance the efficiency of endothelial cell isolation and growth. Thus, the use of specific endothelial cell mitogen such as VEGF, can shorten the time period from cell harvesting to seeding enabling lower initial graft seeding densities to be used.

Endothelial cell proliferation and migration is initiated by activation of VEGF receptors (16). Although three VEGF receptors have been identified and characterized, it is the *KDR* receptor that plays a key role in the angiogenic switch that is comprised of proliferation and migration
 5 (16,17).

VEGF offers distinct advantages over other less endothelial cell-specific growth factors that might enhance graft endothelialization since it has reduced impact on other vascular cells, in particular smooth muscle cells, and as such reduces the potential for adverse stimulatory effects.
 10 Thus, VEGF released by genetically modified cells seeded onto artificial graft will recruit endothelial cells, but not smooth muscle cells, from anastomosis sites.

Although genetically transfected endothelial cells over-expressing VEGF are capable of enhanced growth as compared to non-transformed

endothelial cells, artificial vascular grafts incorporating such transformed cells are functionally limited since the transformed cells coating the luminal surface of the graft are incapable of withstanding flow forces following implantation.

5 There is thus a widely recognized need for and it would be highly advantageous to have an artificial vascular graft capable of supporting blood flow following implantation and yet devoid of the abovementioned limitations of prior art grafts.

10

SUMMARY OF THE INVENTION

According to one aspect of the present invention there is provided an artificial vascular graft comprising a synthetic tubular element having a luminal surface being coated with a plurality of endothelial cells being

genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

According to another aspect of the present invention there is provided a method of replacing or bypassing at least a portion of a vascular system of an individual, the method comprising the step of
5 implanting into the vascular system of the individual an artificial vascular graft, so as to form a fluid communication between the vascular system and the artificial vascular graft, the artificial vascular graft including a synthetic tubular element having a luminal surface being coated with a
10 plurality of endothelial cells being genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

According to yet another aspect of the present invention there is provided a method of producing an artificial vascular graft, the method

comprising the steps of: (a) genetically transforming endothelial cells to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor; and (b) seeding and culturing the endothelial cells within a synthetic tubular element having a luminal surface until sufficient endothelialization of the luminal surface is achieved.

According to further features in preferred embodiments of the invention described below, a first portion of the plurality of endothelial cells is genetically transformed to express the at least one endothelial cell proliferating growth factor and further wherein a second portion of the plurality of endothelial cells is genetically transformed to express the at least one cellular adherence factor.

According to still further features in the described preferred embodiments the luminal surface is of a substance selected from the group

consisting of polytetrafluoroethylene (PTFE), expanded polytetrafluoroethylene (ePTFE), polyester fibers, Dacron and processed animal blood vessels. According to still further features in the described preferred embodiments the synthetic tubular element is of an inner cross sectional area which is substantially equivalent to an inner cross sectional area of a blood vessel.

According to still further features in the described preferred embodiments the inner cross sectional area of the synthetic tubular element is within a range of about 7 to 700 mm² According to still further features in the described preferred embodiments the plurality of endothelial cells are derived from a source selected from the group consisting of a segment of a vein, bone marrow, peripheral blood progenitor cells and circulating endothelial cells.

According to still further features in the described preferred
embodiments the plurality of endothelial cells are derived from the
recipient of the artificial vascular graft.

According to still further features in the described preferred
5 embodiments the plurality of endothelial cells are derived from embryonic
or mature tissues of a human or an animal donor.

According to still further features in the described preferred
embodiments the plurality of endothelial cells form a confluent monolayer
at the inner luminal surface.

10 According to still further features in the described preferred
embodiments at least one endothelial cell proliferating growth factor is
selected from the group consisting of VEGF, basic or acidic FGF, HGF and
any other endothelial growth factor.

According to still further features in the described preferred
embodiments at least one cellular adherence factor is DANCE, UP50 or any
other molecule capable of promoting cellular adherence to graft material.

According to still further features in the described preferred
5 embodiments the plurality of endothelial cells are further genetically
transformed to express at least one marker polypeptide.

According to still another aspect of the present invention there is
provided a nucleic acid expression construct comprising: (a) a first
polynucleotide segment encoding an endothelial cell proliferating growth
factor; and (b) a second polynucleotide segment encoding cellular
10 adherence factor.

According to still another aspect of the present invention there is
provided a nucleic acid expression construct system comprising: (a) a first
nucleic acid expression construct including a first polynucleotide segment

encoding an endothelial cell proliferating growth factor; and (b) a second nucleic acid expression construct including a second polynucleotide segment encoding cellular adherence factor.

According to still further features in the described preferred
5 embodiments the first and the second nucleic acid expression constructs further including at least one additional polynucleotide segment encoding a marker polypeptide.

According to still another aspect of the present invention there is provided a kit comprising a stand for engaging at least one tube, the at
10 least one tube including a nucleic acid expression constructs system, the nucleic acid expression construct system including: (a) a first nucleic acid expression construct including a first polynucleotide encoding an endothelial cell proliferating growth factor; and (b) a second nucleic acid expression construct including a second polynucleotide encoding cellular

adherence factor.

According to still another aspect of the present invention there is provided a kit comprising a stand for engaging at least one tube, the at least one tube including a nucleic acid expression construct, the nucleic acid expression construct including: (a) a first polynucleotide encoding an endothelial cell proliferating growth factor; and (b) a second polynucleotide encoding cellular adherence factor.

According to still another aspect of the present invention there is provided method of producing genetically transformed endothelial cells, the method comprising the steps of: (a) obtaining endothelial cells from a source selected from the group consisting of a segment of a vein, bone marrow progenitor cells, peripheral blood stem cells and circulating endothelial cells; and (b) transforming the endothelial cells to express at least one endothelial cell proliferating growth factor and at least one

cellular adherence factor concurrently or separately.

According to still further features in the described preferred
embodiments the nucleic acid expression construct further comprising at
least one promoter sequence being for directing the expression of at least
5 one of the first and the second polynucleotide segments.

According to still further features in the described preferred
embodiments the first polynucleotide segment is transcriptionally linked to
the second polynucleotide segment whereas the first and the second
polynucleotide segment are under the transcriptional control of a single
10 promoter sequence of the at least one promoter sequence.

According to still further features in the described preferred
embodiments the nucleic acid construct further comprising a linker
sequence being interposed between the first and the second
polynucleotide segments.

According to still further features in the described preferred embodiments the linker sequence is selected from the group consisting of IRES and a protease cleavage recognition site.

According to still further features in the described preferred
5 embodiments at least one promoter is functional in eukaryotic cells.

According to still further features in the described preferred
embodiments at least one promoter is selected from the group consisting
of a constitutive promoter, an inducible promoter and a tissue specific
promoter.

10 According to still further features in the described preferred
embodiments nucleic acid expression construct further comprising: (c) a
first promoter sequence being for directing the expression of the first
polynucleotide segment; and (d) a second promoter sequence being for
directing the expression of the second polynucleotide segment.

According to still further features in the described preferred embodiments the first promoter and the second promoter are selected from the group consisting of a constitutive promoter, an inducible promoter and a tissue specific promoter.

5 According to still further features in the described preferred embodiments the inducible promoter are regulatable by same effector molecule.

According to still further features in the described preferred embodiments the nucleic acid expression construct, further comprising at
10 least one additional polynucleotide segment encoding a marker polypeptide.

According to still further features in the described preferred embodiments the marker polypeptide is selected from the group consisting of a selection polypeptide and a reporter polypeptide.

According to still further features in the described preferred embodiments at least one additional polynucleotide segment is transcriptionally linked to the at least one of the first and the second polynucleotide segments.

- 5 According to still further features in the described preferred embodiments at least one additional polynucleotide segment is transcriptionally linked to the at least one of the first and the second polynucleotide segments via linker segment.

- According to still further features in the described preferred
10 embodiments the linker sequence is selected from the group consisting of IRES and a protease cleavage recognition site.

 According to still further features in the described preferred embodiments at least one additional polynucleotide segment is translationally fused to at least one of the first and the second

polynucleotide segments.

According to still another aspect of the present invention there is provided a transformed endothelial cell being genetically transformed to express at least one endothelial cell proliferating growth factor and at least
5 one cellular adherence factor.

According to still further features in the described preferred
embodiments the transformed endothelial cell further genetically transformed to express at least one marker polypeptide.

According to still further features in the described preferred
10 embodiments the endothelial cell is derived from a source selected from the group consisting of a segment of a vein, bone marrow, peripheral blood progenitor cells and circulating endothelial cells.

According to still further features in the described preferred
embodiments the endothelial cell is derived from a source selected from

the group consisting of embryonic or mature tissues of a human donor and an animal source.

According to still further features in the described preferred embodiments the cellular proliferating growth factor is selected from the group consisting of VEGF, acidic and basic FGF, HGF and any other endothelial cell growth factor. According to still further features in the described preferred embodiments the cellular adherence factor is DANCE, UP50 or any other molecule capable of promoting cellular adherence to graft material.

10 The present invention successfully addresses the shortcomings of the presently known configurations by providing an artificial vascular graft and method of generating same, which graft is useful for vascular replacement or vascular bypass.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention is herein described, by way of example only, with reference to the accompanying drawings. With specific reference now to

5 the drawings in detail, it is stressed that the particulars shown are by way of example and for purposes of illustrative discussion of the preferred embodiments of the present invention only, and are presented in the cause of providing what is believed to be the most useful and readily understood description of the principles and conceptual aspects of the invention. In

10 this regard, no attempt is made to show structural details of the invention in more detail than is necessary for a fundamental understanding of the invention, the description taken with the drawings making apparent to those skilled in the art how the several forms of the invention may be embodied in practice.

In the drawings:

FIG. 1 is a perspective view of an artificial vascular graft of the present invention.

FIGs. 2a-c illustrate VEGF expression vector constructs according to the teachings of the present invention.

FIGs. 3a-c illustrates a Urine p50 protein (UP50) or a neural crest epidermal growth factor-like (DANCE) expression vector construct according to the teachings of the present invention.

FIG. 4a illustrates a PTFE (polytetrafluoroethylene) graft seeded with human endothelial cells. The cells were infected with recombinant adenoviral vector encoding nuclear targeted LacZ gene. Blue staining indicates an infected cell expressing the transgene. Staining was performed 24 hours following infection.

FIG. 4b illustrates primary human endothelial cells seeded on a PTFE graft and infected with recombinant adenoviral vector co-expressing the VEGF and green fluorescent protein (GFP) genes. Green cells co-express VEGF and GFP.

5 FIG. 5a illustrates a PTFE graft seeded with primary human and sheep venous endothelial cells following transformation with recombinant pseudo-typed retroviral vector encoding the LacZ gene. Blue staining indicates an infected cell expressing the transgene.

10 FIG. 5b illustrates primary sheep endothelial cells seeded on a PTFE graft following transformation with recombinant pseudo-typed retroviral vector co-expressing VEGF and GFP. Green cells indicate cells co-expressing VEGF and GFP.

FIGs. 6a-b illustrate binding and cross linking of ^{125}I -VEGF to specific receptors expressed on human saphenous vein endothelial cells

(HSVEC). VEGF receptor expression in EC was detected at day 2 (Figure 6a) and day 5 (Figure 6b) following adenoviral infection. Cells were washed extensively and incubated with ^{125}I -VEGF (1 ng/ml or 5 ng/ml). Binding specificity was detected by competition in the presence of 1.5 μ g/ml unlabeled recombinant VEGF₁₆₅ (marked as + lanes at the top of Figure 6a). The cells were lysed and ^{125}I -VEGF -receptor complexes were separated on 6% polyacrylamide gel and exposed to XAR film for autoradiography. The lanes in Figures 6a-b are as follows: 1. control uninfected cells; 2. cells infected with rAdlacZ; 3. cells infected with rAdVEGF; 4. uninfected EC supplemented with recombinant VEGF₁₆₅ (2 ng/ml); 5. uninfected EC supplemented with recombinant VEGF₁₆₅ (10 ng/ml); and 6. uninfected EC supplemented with conditioned medium collected from rAdVEGF infected cells.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is of an artificial vascular graft and methods of generating same which can be utilized to replace or bypass damaged
5 and/or occluded blood vessels. Specifically, the present invention is of a method of generating synthetic vascular grafts coated with mammalian endothelial cells which grafts can provide a viable alternative to presently utilized natural and synthetic vascular grafts.

The principles and operation of the present invention may be better
10 understood with reference to the drawings and accompanying descriptions.

Before explaining at least one embodiment of the invention in detail, it is to be understood that the invention is not limited in its application to the details of construction and the arrangement of the components set

forth in the following description or illustrated in the drawings. The invention is capable of other embodiments or of being practiced or carried out in various ways. Also, it is to be understood that the phraseology and terminology employed herein is for the purpose of description and should

5 not be regarded as limiting.

Artificial grafts are yet to be extensively utilized in surgical bypass procedures. One of the limitations plaguing presently available artificial vascular grafts as well as some natural vascular grafts is the lack of durability over extended periods of time. Although the use of endothelial

10 cells greatly enhances the quality of coated artificial grafts, by improving performance in general and by improving thrombolytic activity in particular, such grafts typically suffer from loss of cell coating under blood flow forces since the coated cells cannot withstand the forces exerted thereupon by the flow of blood.

Thus, the present inventors have suggested that a genetically modified population of endothelial cells which co-express a cellular adherence factor, such as DANCE or UP50 along with a cellular growth factor, such as VEGF, or a mixture of two subpopulations of genetically modified endothelial cells, one expressing a cellular adherence factor and the other an endothelial cell growth factor are advantageous for use in artificial vascular grafts since the presence of a cellular adherence factor in these cells greatly increases the adherence to the luminal surface of the artificial graft substrate which greatly increases the durability of these vascular grafts. The proteins expressed and secreted by such cells will lead to the complete coverage of the grafts with endothelial cells which are capable of withstanding the forces exerted by the flow of blood therein.

Thus according to one aspect of the present invention and as specifically shown in Figure 1, there is provided an artificial vascular graft

which is referred to hereinunder as graft **10**.

Graft **10** includes a synthetic tubular element **12** having a luminal surface **14**. Preferably, synthetic tubular element **12** is of an inner cross sectional area of about 7 to 700 mm² or any cross sectional area or diameter
 5 which is substantially equivalent to an inner cross sectional area or diameter of a blood vessel.

Luminal surface **14** is fabricated from a substance including but are not limited to, polytetrafluoroethylene (PTFE), expanded polytetrafluoroethylene (ePTFE), polyester fibers, Dacron™ or processed
 10 blood vessels derived from an animal or a human.

As is further described hereinunder and in the Examples section which follows, luminal surface **14** is coated with a plurality of endothelial cells **16** which are genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular

adherence factor. As such luminal surface **14** preferably includes structures such as pits or projection which facilitate adherence of endothelial cells **16** to surface **14**.

Graft **10** is fabricated by genetically transforming a population of
5 endothelial cells **16** with at least one endothelial cell proliferating growth factor and at least one cellular adherence factor and culturing the transformed cells within element **12** until sufficient endothelialization of luminal surface **14** is achieved.

It will be appreciated that the population of endothelial cells **16** can
10 be transformed with both the endothelial cell proliferating growth factor and the cellular adherence factor, or alternatively a first portion of such population can be transformed with the endothelial cell proliferating growth factor, while a second portion of this population is transformed with cellular adherence factor.

As is further described in the examples section which follows
endothelial cells **16** are cultured with element **12** in a manner which is
conducive for the formation of a confluent monolayer upon luminal
surface **14**.

5 According to a preferred embodiment of the present invention,
endothelial cells **16** are prepared from, for example, a segment of a vein,
bone marrow progenitor cells, peripheral blood stem cells or circulating
endothelial cells of an intended recipient of graft **10** or a syngeneic donor.

Alternatively xenogeneic tissue can also be utilized for the
10 preparation of endothelial cells **16** providing measures are taken prior to,
or following fabrication of graft **10**, so as to avoid cell rejection.
Numerous methods for preventing or alleviating rejection are known in the
art and as such no further detail is given herein.

As is further described in the Examples section below, the co-expression of a cellular adherence factor along with a cellular growth factor enables endothelial cells **16** of graft **10** of the present invention to withstand flow forces which would normally lead to endothelial cell loss

5 from the luminal surface of prior art artificial vascular grafts.

Thus, graft **10** of the present invention can be successfully utilized in vascular surgical procedures aimed at replacing or bypassing a damaged or occluded blood vessel.

Thus, according to another aspect of the present invention there is

10 provided a method of replacing or bypassing at least a portion of a vascular system of an individual. The method is effected by implanting into the vascular system of the individual graft **10**, so as to form a fluid communication between the vascular system and graft **10**.

As used herein the phrase “genetic transformation” refers to the introduction of exogenous polynucleotide sequences into a cell. Such sequences can integrate into the genome of the cell, or alternatively, such exogenous sequences can exist in the nucleus or cytoplasm of the cell in a transient manner.

The transformed endothelial cells according to the teachings of the present invention express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor. Such factors can be expressed from exogenous polynucleotide sequences introduced into the endothelial cells, or alternatively such factors can be endogenous factors naturally expressed by the cell in which case, the cell is transformed with sequences directed at causing the over expression of the endogenous factors.

As used herein, the term “overexpression” refers to expression levels which exceeded those normally found in a cell. Over expression can be effected by introducing additional copies of an endogenous gene into a cell thus increasing the copy number of the endogenous gene or genes expressed in a cell, or by introducing enhancer sequence elements via gene knock-in procedures, which elements upregulate the transcription or translation of the endogenous genes.

The exogenous polynucleotide sequences are preferably included in one or more nucleic acid constructs which are used to transform endothelial cells.

Thus according to another aspect of the present invention there is provided a nucleic acid expression construct useful for generating endothelial cells **16** of the present invention. The construct includes a first polynucleotide segment encoding an endothelial cell proliferating

growth factor and a second polynucleotide segment encoding a cellular adherence factor.

According to preferred embodiments of the present invention, the endothelial cell proliferating growth factor is, for example, VEGF, acidic or basic FGF or HGF, while the cellular adherence factor is, for example, DANCE, UP50 or any other molecule capable of promoting endothelial cell adherence to the graft material. Such factors can be encoded by sequences derived from human cells or any other mammalian cells provided the expressed factors are functional in endothelial cells.

The construct according to this aspect of the present invention also includes one or more promoter sequences for directing the expression of the first and second polynucleotide segments in endothelial cells. Such a promoter can be any mammalian functional promoter which is either constitutive, tissue specific or inducible.

As is further described in the examples section herein below, the endothelial cell proliferating growth factor and the cellular adherence factor are preferably expressed in a different temporal pattern.

As such the construct includes two promoter sequences each
5 being for directing the expression of a single polynucleotide segment.

Preferably such promoters are inducible promoters regulatable by the same effector molecule. Preferably, the promoter sequences are selected such that one promoter is upregulatable by an effector molecule, while the second promoter is downregulatable by the same effector.

10 Examples of suitable inducible promoters and effectors are given in the Examples section hereinbelow.

Notwithstanding from the above, it will be appreciated that a single promoter sequence can also be utilized by the construct of the present invention provided the first and second polynucleotide segments are

transcriptionally linked and provided that an internal ribosome entry site (IRES) is included for directing the translation of the second segment of the polycistronic message transcribed.

The two polynucleotide segments of the construct according to
5 this aspect of the present invention can also be translationally fused provided a protease cleavage site is provided therebetween which allows cleavage and separation of the two polypeptides in endothelial cells.

It will further be appreciated that each of the two polynucleotide segments can be included in a separate nucleic acid constructs, which can
10 be co-introduced into endothelial cells.

Thus according to yet another aspect of the present invention there is provided a nucleic acid expression construct system useful in generating endothelial cells **16** of the present invention. The construct system of this aspect of the present invention includes a first nucleic acid

expression construct including a first polynucleotide segment encoding an endothelial cell proliferating growth factor and a second nucleic acid expression construct including a second polynucleotide segment encoding cellular adherence factor.

5 The construct or construct system of the present invention preferably also includes additional polynucleotides segments encoding marker genes, selection marker genes and the like, so as to enable selection of transformed cells and/or monitoring of the expression of the endothelial cell proliferating growth factor and/or the cellular adherence
10 factor. Examples of suitable reporter genes include, but are not limited to, beta-galactosidase, luciferase, GFP and the like. Examples of selection markers include antibiotic resistance genes and the like.

It will be appreciated that to monitor expression of the endothelial cell proliferating growth factor and/or the cellular adherence factor, the

reporter gene can be provided in transcriptional or translational fusion to the polynucleotide segment encoding the factor or it can be placed under a transcriptional control of a promoter sequence identical to that directing the transcription of the factor.

5 It will be appreciated that the polynucleotide segments encoding the endothelial cell proliferating growth factor and/or the cellular adherence factor, can be ligated into a commercially available expression vector system suitable for transforming mammalian cells and for directing the expression of these factors within the transformed cells. It will be
10 appreciated that such commercially available vector systems can easily be modified via commonly used recombinant techniques in order to replace, duplicate or mutate existing promoter or enhancer sequences and/or introduce any additional polynucleotide sequences.

Suitable mammalian expression vectors include, but are not limited to, pcDNA3, pcDNA3.1(+/-), pZeoSV2(+/-), pSecTag2, pDisplay, pEF/myc/cyto, pCMV/myc/cyto, pCR3.1, which are available from Invitrogen, pCI which is available from Promega, pBK-RSV and pBK-CMV
5 which are available from Stratagene, pTRES which is available from Clontech, and their derivatives.

As mentioned hereinabove, the endothelial cells of the present invention can also include exogenous polynucleotide sequences directed at over expressing endogenously encoded factors.

10 Thus, according to yet another aspect of the present invention, there is provided a nucleic acid construct or construct system which serves to upregulate expression of endogenously encoded factors. Such a construct or constructs can include transcriptional regulatory sequences which when provided in cis to endogenous sequences encoding the

endothelial cell proliferating growth factor and/or the cellular adherence factor, can upregulate the transcription thereof.

Alternatively, such a construct or constructs can encode translational regulatory sequences which when provided in trans to
 5 endogenous sequences encoding the endothelial cell proliferating growth factor and/or the cellular adherence factor, can upregulate the translation thereof.

Standard gene knock-in techniques can be utilized to introduce cis acting transcriptional regulatory sequences into a genome of the
 10 endothelial cells. For a review of gene knock-in methodology see, for example, United States Patent Nos. 5,487,992, 5,464,764, 5,387,742, 5,360,735, 5,347,075, 5,298,422, 5,288,846, 5,221,778, 5,175,385, 5,175,384, 5,175,383, 4,736,866 as well as Burke and Olson, Methods in Enzymology, 194:251-270, 1991; Capecchi, Science 244:1288-1292, 1989; Davies *et al.*,

Nucleic Acids Research, 20 (11) 2693-2698, 1992; Dickinson *et al.*, Human Molecular Genetics, 2(8):1299-1302, 1993; Duff and Lincoln, "Insertion of a pathogenic mutation into a yeast artificial chromosome containing the human APP gene and expression in ES cells", Research Advances in

5 Alzheimer's Disease and Related Disorders, 1995; Huxley *et al.*, Genomics, 9:742-750 1991; Jakobovits *et al.*, Nature, 362:255-261 1993; Lamb *et al.*, Nature Genetics, 5: 22-29, 1993; Pearson and Choi, Proc. Natl. Acad. Sci. USA, 1993, 90:10578-82; Rothstein, Methods in Enzymology, 194:281-301, 1991; Schedl *et al.*, Nature, 362: 258-261, 1993; Strauss *et al.*, Science,

10 259:1904-1907, 1993, WO 94/23049, WO93/14200, WO 94/06908 and WO 94/28123 also provide information.

The nucleic acid constructs of the present invention can be introduced into endothelial cells via any standard mammalian transformation method. Such methods include, but are not limited to, direct

DNA uptake techniques, and virus or liposome mediated transformation
(for further detail see, for example, "Methods in Enzymology" Vol. 1-317,
Academic Press).

Thus, the present invention provides an improved artificial vascular
5 graft which is internally coated with genetically transformed endothelial
cells expressing a cellular growth factor and a cellular adhesion factor.
Expression of the cellular adherence factor, in addition to the cellular
growth factor, enables the endothelial cells of the coating to adhere with
greater force to the luminal surface of the artificial vascular graft thereby
10 providing a durable cellular coating which can withstand, for extended
periods of time, the flow forces exerted upon the luminal surface of the
artificial vascular graft following implantation.

Additional objects, advantages, and novel features of the present invention will become apparent to one ordinarily skilled in the art upon examination of the following examples, which are not intended to be limiting. Additionally, each of the various embodiments and aspects of the present invention as delineated hereinabove and as claimed in the claims section below finds experimental support in the following examples.

EXAMPLES

Reference is now made to the following examples, which together with the above descriptions, illustrate the invention in a non limiting fashion.

Generally, the nomenclature used herein and the laboratory procedures utilized in the present invention include molecular, biochemical, microbiological and recombinant DNA techniques. Such

techniques are thoroughly explained in the literature. See, for example,

"Molecular Cloning: A laboratory Manual" Sambrook et al., (1989);

"Current Protocols in Molecular Biology" Volumes I-III Ausubel, R. M.,

ed. (1994); Ausubel et al., "Current Protocols in Molecular Biology", John

5 Wiley and Sons, Baltimore, Maryland (1989); Perbal, "A Practical Guide to

Molecular Cloning", John Wiley & Sons, New York (1988); Watson et al.,

"Recombinant DNA", Scientific American Books, New York; Birren et al.

(eds) "Genome Analysis: A Laboratory Manual Series", Vols. 1-4, Cold

Spring Harbor Laboratory Press, New York (1998); methodologies as set

10 forth in U.S. Pat. Nos. 4,666,828; 4,683,202; 4,801,531; 5,192,659 and

5,272,057; "Cell Biology: A Laboratory Handbook", Volumes I-III Cellis, J.

E., ed. (1994); "Culture of Animal Cells - A Manual of Basic Technique" by

Freshney, Wiley-Liss, N. Y. (1994), Third Edition; "Current Protocols in

Immunology" Volumes I-III Coligan J. E., ed. (1994); Stites et al. (eds),

"Basic and Clinical Immunology" (8th Edition), Appleton & Lange, Norwalk, CT (1994); Mishell and Shiigi (eds), "Selected Methods in Cellular Immunology", W. H. Freeman and Co., New York (1980); available immunoassays are extensively described in the patent and scientific literature, see, for example, U.S. Pat. Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; 4,098,876; 4,879,219; 5,011,771 and 5,281,521; "Oligonucleotide Synthesis" Gait, M. J., ed. (1984); "Nucleic Acid Hybridization" Hames, B. D., and Higgins S. J., eds. (1985); "Transcription and Translation" Hames, B. D., and Higgins S. J., eds. (1984); "Animal Cell Culture" Freshney, R. I., ed. (1986); "Immobilized Cells and Enzymes" IRL Press, (1986); "A Practical Guide to Molecular Cloning" Perbal, B., (1984) and "Methods in Enzymology" Vol. 1-317, Academic Press; "PCR Protocols: A Guide To Methods And Applications", Academic Press, San

Diego, CA (1990); Marshak et al., "Strategies for Protein Purification and Characterization - A Laboratory Course Manual" CSHL Press (1996); all of which are incorporated by reference as if fully set forth herein. Other general references are provided throughout this document. The

5 procedures therein are believed to be well known in the art and are provided for the convenience of the reader. All the information contained therein is incorporated herein by reference.

EXAMPLE 1

Since artificial graft coating with endothelial cells is of a great

10 importance in surgical treatment of vascular diseases, a system was developed which enables complete endothelialization of artificial grafts expressing at least one endothelial cell proliferating growth factor and at least one cellular adherence factor. To generate such grafts a population of endothelial cells utilized as graft coating are transformed with several

expression vectors as describe below.

Expression vectors and virus particles

Construction of rAdGFP and rAdVEGF-GFP recombinant

Adenoviral-based vectors:

5 The recombinant adenoviral vectors expressing the human VEGF165 and/or the GFP genes were constructed in two steps. The first step included construction of two shuttle vector plasmids expressing the GFP and the VEGF165 -GFP genes. For this purpose a shuttle vector plasmid containing the left arm (16%) of the Ad5 genome (A kind gift from

10 Shenk T, Princeton University, NJ, USA) with a deletion in the E1 region was used. This deletion was replaced by IRES-GFP fragment (Clontech, Ca, USA) regulated by the CMV5 promoter. This shuttle plasmid was used for construction of rAdGFP. A second shuttle vector was constructed by ligating a 600bp BamHI fragment containing the human VEGF165 cDNA

which includes the secretion signal sequence (Genbank Accession number AB021221) into the BglII site between the CMV5 promoter and the IRES sequence. This bi-cistronic shuttle vector which expresses both the VEGF165 and the GFP genes was used for construction of rAdVEGF-GFP.

5 The second step was effected via homologous recombination between each shuttle vector and the adenoviral plasmid (pJM17 a kind gift from Shenk T, Princeton University, NJ, USA) via co-transfection into the 293 cell line. The pJM17 plasmid contained the adenovirus genome, excluding the E3 region, and an insert (pBRX) in the E1 region of the virus.

10 Homologous recombination between the expression plasmid and pJM17 following transfection into the 293 cells replaced the E1 region and pBRX insert with the expression vector from the shuttle plasmid. Plaque formation occurred between 2 to 4 weeks following co-transfection.

Individual plaques were isolated and viral extracts were amplified by

infecting a 293 cell line. Titer of viral stocks, as determined by plaque assay in these cells, ranged from 10^{10} to 10^{11} pfu/ml. Expression of VEGF was confirmed by western blotting while expression of GFP was confirmed using an inverted fluorescent microscope. Recombinant vectors encoding

5 the nuclear targeted beta-galactosidase (LacZ) (Clontech, CA, USA) were prepared using similar methods.

Preparation of retrovirus vector expressing the VEGF165 -GFP

genes:

Figures 2a-c illustrate the VEGF-GFP expression vector constructs

10 according to the teachings of the present invention. The recombinant retroviral vectors expressing the human VEGF165 and/or the EGFP genes (EGFP -Clontech, Ca, USA) were constructed by separately cloning two different vectors into the LXSNI plasmid (Clontech, Ca, USA). For construction of LXSNI-GFP expressing EGFP alone, an EcoRI-HpaI 1400bp

fragment containing the IRES element and EGFP gene was inserted into the specific restriction sites of the LXSNI plasmid multiple cloning site (MCS). For construction of the LXSNI-VEGF-GFP bi-cistronic plasmid which co-expresses the VEGF165 and the EGFP genes, a 2.0 kB EcoRI-MunI fragment containing VEGF165 (600bp), and the IRES and EGFP sequences were ligated into the EcoRI restriction site of the LXSNI MCS. Expression from both vectors is regulated by the MoMULV long terminal repeat (LTR) (Clontech, Ca, USA). For retroviral vectors production, 10 µg of LXSNI-GFP or LXSNI-VEGF-GFP plasmid DNA were transfected into 293E3 ecotopic packaging cells and incubated for 48 hours, following which the supernatant from confluent cultures of G418 resistant producer cells were collected, filtered (0.45 µm) and added to PA317 amphotropic packaging cells. The transduced cells were exposed to G418 selection and the supernatant was collected again after 48 hours and used to infect

TEFLYGA amphotropic packaging cells which uniquely express the GALV envelope for high transduction efficiency of endothelial cells. This packaging cell line has the additional advantage of producing amphotropic particles resistant to human serum inactivation. Following G418 selection of transduced TEFLYGA cells, individual colonies were collected and screened for EGFP and VEGF165 expression. Viral titers of each colony were determined by TE671 cells transduction. Titters ranged between 10^5 and 10^6 pfu/ml. The highest-titer producing colonies were selected and their supernatants were collected and stored at -80 °C.

The above described steps can also be applied to the construction of a cellular adherence factor expression vector. For example, a PCR amplified fragment of human derived UP50 nucleotide sequence (Genbank Accesssion number AF093118) or DANCE nucleotide sequence (GenBank Accession Number AF112152) can be ligated into a mammalian expression

vector suitable for expression of the cellular adherence factor in endothelial cells (Figures 3a-b).

Transformed cell preparation

Autologous endothelial cell harvesting:

5 Endothelial cells are harvested from 5cm long venous segments of hind limb. Cells are harvested by collagenase digestion as previously described (Flugelman MY, Virmani R, Leon MB, Bowman RL, Dichek DA. Circulation Research 1992;70:348-354). Cells from passages 3-9 are collected to ensure stability of cell characteristics. Cell identity is tested
10 with immunohistochemistry for von Willebrand's factor.

Bone marrow progenitor cell infection with VEGF encoding

vectors:

Bone marrow progenitor cells were infected with recombinant adenoviral vectors encoding VEGF-GFP. Differentiation is tested in

infected cells using morphology and immunohistochemistry for von Willebrand's factor. For more details of cell harvesting see: Asahara T, Takahashi T, Masuda H, Kalka C, Chen D, Iwaguro H, Inai Y, Silver M, Isner JM. EMBO J 1999;18:3964-72, and Huang XL, Takakura N, Suda T.

5 Biochem Biophys Res Commun 1999;264:133-8.

Regulation of gene expression after seeding:

An effector regulatable expression system can be used in order to regulate gene expression of VEGF and DANCE in endothelial cells. For example, VEGF expression can be downregulated via tetracycline, while

10 DANCE expression is upregulated thereby. With these systems, the seeded cells can be made to express VEGF in the first week when cell proliferation is needed, while use of tetracycline one week following surgery downregulates VEGF expression and upregulates DANCE expression to increase cell adherence to the grafts. (Agha-Mohammadi S,

Lotze MT. Regulatable systems: applications in gene therapy and replicating viruses. J Clinical Investigations 2000;105:1177-1183.)

Transduction of human and sheep saphenous vein endothelial

cells by retroviral vectors:

5 Endothelial cells were seeded (105 cells/35 mm dish) in fibronectin coated plates and grown in M199 containing 20% fetal serum. After 24 hours the cells were exposed to the cationic polymer DEAE-dextran (0.1µg/ml) for 1 hour before transduction, washed 3 times with PBS and than transduced with high titer virus. Following incubation for 4 hours at 10 37 °C, the virus is replaced by fresh medium containing 20% fetal serum. A day later the cells are trypsinized and seeded again on fibronectin coated PTFE (approximately 1 cm²). Figure 4a-6b illustrate the expression levels from the various constructs as indicated by LacZ or GFP activity.

Analysis of KDR binding expression and activation:

Iodination of 5µg of human recombinant VEGF165 is performed by the chloramine T method. Free Iodine is separated from 125I-VEGF165 using heparin-Sepharose affinity chromatography. The specific activity of 125I-VEGF165 obtained is usually 0.5-1.5 x 10⁵ cpm/ng. For more details see: Vaisman N, Gospodarowicz D, Neufeld G. J Biol Chem 1990;265:19461-19466.

The grafts are cut longitudinally and placed in a cell culture dish and then washed in cold PBS. The binding of 125I-VEGF165 to the cell surface receptors is carried out in binding buffer for 2 hours at 4 °C followed by cross linking in the presence of disuccinimidyl substrate. Cells seeded on the grafts are lysed by lysis buffer and the VEGF-receptor complexes are separated on SDS-polyacrylamide gel and analyzed by autoradiography. The gels are stained by Coomassie-blue to assess the

amounts of loaded proteins. Immunoprecipitation for the phosphorylated and unphosphorylated fractions of KDR is used for the evaluation of KDR activation. Results of the binding assay are shown in Figures 6a-b. Table 1 represents the KDR and the neuropilins bands intensity at day 5 following infection (5 ng/ml 125 I-VEGF treated cells). Numbers represent relative densitometric values following standardization of the protein quantity loaded on the gel.

Table 1

Lanes:	1	2	3	4	5	6
bands:	Control	rAdLacZ	rAdVEGF	VEGF ₁₆₅ 2 ng/ml	VEGF ₁₆₅ 10 ng/ml	rAdVEGF Supernatant
KDR:	1	0.85	6	0.9	7.2	6.4
Neuropilins	1	0.77	5.2	1.3	2.5	4.3

10

Analysis of recombinant protein expression:

VEGF is measured in supernatant collected from tissue culture dishes containing the seeded grafts. ELISA is used to measure VEGF production over 24 hours period. Similar methods will be used for the

DANCE protein.

EXAMPLE 2

Synthetic Grafts seeding techniques

Grafts seeding techniques are detailed in: Tseng, D.Y. and ER

5 Edelman, J Biomed Mater Res 1998;42:188-98. A seeding apparatus is

used to allow simultaneous seeding of several grafts. A suitable seeding

apparatus is described in: Mazzucotelli JP, Roudiere JL, Bernex F, Bertrand

P, Leandri J, Loisanse D., Artif Organs 1993;17:787-90.

EXAMPLE 3

Flow Apparatus

10

A suitable flow apparatus for use with the present invention is described in: Flugelman MY, Virmani R, Leon MB, Bowman RL, Dichek DA. Circulation Research 1992;70:348-354. To be able to calculate the shear forces operating on the grafts some modifications from the original

flow apparatus are performed in accordance with: Schneider A, Chandra M, Lazarovici G et al. Thromb Haemost 1997;78:1392-8.

Endothelial cell quantitative morphometry

At the end of the incubation period the grafts are cut longitudinally,
 5 examined under fluorescent microscope, photographed and the images
 processed by an image analysis system.

t-PA and Urokinase measurements

t-PA level and activity are measured from supernatant collected
 from seeded grafts. t-PA and urokinase are tested by ELISA while t-PA
 10 activity is measured as described in Dichek DA, Nussbaum O, Degen SJF,
 Anderson WF, Blood 1991;77:533-41.

Cell recovery from grafts

Cells are recovered from the PTFE grafts by EDTA-trypsin
 digestion. Cells are counted following digestion and re-plated in a tissue

culture dish precoated with fibronectin to assess viability. A time from plating to full confluence is recorded. In cells transduced with pseudo-retroviral vectors VEGF and DANCE expression is measured.

EXAMPLE 4

5

Animal experiments

Short venous segment of sheep hind limb are used for endothelial cell harvesting. Cells are expanded in the laboratory, gene transfer with viral vectors is performed and the seeded grafts are implanted end-to-end proximally and distally into the carotid arteries of the donor sheep. A graft
10 seeded with genetically modified cells is implanted in one side of a carotid artery, while a graft seeded with cells expressing only a marker gene is implanted in the other side of the carotid artery. Short-term and long-term experiments (3, 7, 30 days) are conducted to allow detection of cell adherence and local thrombosis and inflammation. Long term graft

durability and flow will be assessed at 7, 30, and 180 days (n=5 for VEGF and n=5 for DANCE) by external measurements of flow using a Doppler flow meter (Transonic Animal Research Flowmeter, NY, USA). At the end of the follow up period, the grafts are retrieved and neointimal formation is assessed by morphometric measurements of cross sections while seeded cell adherence is tested by detection of marker gene expression.

Although the invention has been described in conjunction with specific embodiments thereof, it is evident that many alternatives, modifications and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all such alternatives, modifications and variations that fall within the spirit and broad scope of the appended claims. All publications, patents, patent applications and sequences disclosed therein and/or identified by a Genbank accession number

mentioned in this specification are herein incorporated in their entirety by reference into the specification, to the same extent as if each individual publication, patent, patent application or sequence was specifically and individually indicated to be incorporated herein by reference. In addition,

5 citation or identification of any reference in this application shall not be construed as an admission that such reference is available as prior art to the present invention.

REFERENCES CITED

1. VA Study Group 141. Comparative evaluation of prosthetic, reversed, and in situ vein bypass grafts in distal popliteal and tibial-peroneal revascularization. Arch Surg 1988;123:434-8.
- 5 2. Londrey GL, Ramsey DE, Hodgson KJ, Barkmeier LD, Sumer DS. Infrapopliteal bypass for severe ischemia: comparison of autogenous vein, composite, and prosthetic grafts. J Vasc J 1991;13:631-6.
3. Darling RC, Linton RR. Durability of femoropopliteal reconstructions. Am J Surgery 1972;123:472-79.
- 10 4. Rutherford RB, Nishikimi N. Graft thrombosis and thromboembolic complications. In Rutherford RB ed. Vascular Surgery. Philadelphia, Pa: WB Saunders Co;1989:501-510.
5. Eickhoff JH, Broome A, Ericsson BF, et al. Four years results of a prospective, randomized clinical trial comparing PTFE and modified human

umbilical for below knee femoropopliteal bypass. J Vasc Surg 1987;6:506-

511.

6. Meinhart J, Deutsch M, Zilla P. Eight years of clinical endothelial cell transplantation. Closing the gap between prosthetic grafts and vein

5 grafts. ASAIO 1999;43:M515-21.

7. Falk J, Townsend LE, Vogel LM, Boyer M, et al. Improved adherence of genetically modified endothelial cells to small diameter expanded PTFE grafts in canine model. J Vasc Surg 1998;27:902-8.

8. Magometschnigg H, Kadletz M, Vodrazka M, et al. Prospective
10 clinical study with in vitro endothelial cell lining of expanded polytetrafluoroethylene grafts in crural repeat reconstruction. J Vasc Surg 1992;15:527-35.

9. Noishiki Y, Tomizawa Y, Yamane Y, Matsumoto A. Autocrine angiogenic vascular prosthesis with bone marrow transplantation. Nature

Medicine 1996;2:90-93.

10. Clowes AW. Improving the interface between biomaterials and the blood - the gene therapy approach. *Circulation* 1996;93:1319-1320.

11. Dichek DA, Anderson J, Kelly AB, Hanson SR, Harker LA.

5 Enhanced in vivo antithrombotic effects of endothelial cells expressing recombinant plasminogen activators transduced with retroviral vectors. *Circulation* 1996;93:301-309.

12. Gillis-Haegerstrand C, Frebelius S, Haegerstrand A, Swedenberg J.

Cultured human endothelial cells seeded on expanded PTFE support

10 thrombin mediated activation of protein C. *J Vasc Surg* 1996;24:226-34.

13. Pasic M, Muller-Glauser W, Odermatt B, Lachat M, Seifert B, Turina

M. Seeding with omental cells prevents late neointimal hyperplasia in small-diameter grafts. *Circulation* 1995;92:2605-2616.

14. Edelman ER. Vascular tissue engineering. *Circ Res* 1999;85:11157.

15. Leung DW, Cachians G, Kuang WJ, Goeddel DV, Ferrara N: Vascular endothelial growth factor is a secreted angiogenic factor. *Science* 1989;246:1306-1309.

5 16. Hanahan D. Signaling vascular morphogenesis and maintenance. *Science* 1997;277:48-50.

17. Vaisman N, Gospodarowicz D, Neufeld G. Characterization of the receptors for vascular endothelial growth factor. *J Biol Chem* 1990;265:19461-19466.

10 18. Sapienza P, di Marzo L, Cucina A, et al. Release of PDGF-BB and bFGF by human endothelial cells seeded on ePTFE vascular grafts. *J Surg Res* 1998;75:24-9.

19. Van Belle E, Tio FO, Couffinhal T, Maillard L, Passeri J, Isner JM. Stent endothelialization. *Circulation* 1997;95:438-448.

20. Folkman J: Therapeutic angiogenesis in ischemic limbs [editorial; comment]. *Circulation* 1998;97:1108-1110

21. Anderson WF. Human gene therapy. *Nature* 392;25-30.

22. Mulligan RC: The basic science of gene therapy. *Science* 260:926-932,
 5 1993.

23. Cosset F-L, Russell SJ. Targeting retrovirus entry. *Gene therapy*
 1996;3:946-956.

24. Nakamura T, Ruiz-Lozano P, Lindner V, et al. DANCE, a novel
 secreted RGD protein expressed in developing, atherosclerotic, and
 10 balloon injured arteries. *JBC* 1999; 274,:22476-22483.

25. Huber TS, Welling TH, Sarkar R, Messina LM, Stanley JC. Effects
 of retroviral-mediated t-PA gene transfer and expression on adherence and
 proliferation of canine endothelial cells seeded onto ePTFE. *J Vasc Surg*
 1995;22:795-803.

26. Dunn PF, Newman KD, Jones M, Yamada I, Shayani V, Virmani R, Dichek DA. Seeding of vascular grafts with genetically modified endothelial cells; secretion of recombinant T-PA results in decreased seeded cell retention in vitro and in vivo. *Circulation* 1996;93:1439-1446.
- 5 27. Zilla P, Greisler HP (Eds). *Tissue engineering of vascular prosthetic grafts*. RG Landes Company, Austin, Texas, USA 1999.

WHAT IS CLAIMED IS:

1. An artificial vascular graft comprising a synthetic tubular element having a luminal surface being coated with a plurality of endothelial cells being genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

2. The artificial vascular graft of claim 1, wherein a first portion of said plurality of endothelial cells is genetically transformed to express said at least one endothelial cell proliferating growth factor and further wherein a second portion of said plurality of endothelial cells is genetically transformed to express said at least one cellular adherence factor.

3. The artificial vascular graft of claim 1, wherein said luminal surface is of a substance selected from the group consisting of polytetrafluoroethylene (PTFE), expanded polytetrafluoroethylene (ePTFE), polyester fibers, Dacron and processed animal blood vessels.

4. The artificial vascular graft of claim 1, wherein said synthetic tubular element is of an inner cross sectional area which is substantially equivalent to an inner cross sectional area of a blood vessel.

5. The artificial vascular graft of claim 4, wherein said inner cross sectional area of said synthetic tubular element is within a range of about 7 to 700 mm².

6. The artificial vascular graft of claim 1, wherein said plurality of endothelial cells are derived from a source selected from the group consisting of a segment of a vein, bone marrow progenitor cells, peripheral blood stem cells and circulating endothelial cells.

7. The artificial vascular graft of claim 1, wherein said plurality of endothelial cells are derived from a recipient of the artificial vascular graft.

8. The artificial vascular graft of claim 1, wherein said plurality of endothelial cells are derived from a human or an animal donor.

9. The artificial vascular graft of claim 1, wherein said plurality of endothelial cells form a confluent monolayer at said inner luminal

surface.

10. The artificial vascular graft of claim 1, wherein said at least one endothelial cell proliferating growth factor is selected from the group consisting of VEGF, acidic or basic FGF and HGF.

11. The artificial vascular graft of claim 1, wherein said at least one cellular adherence factor is DANCE or UP50.

12. The artificial vascular graft of claim 1, wherein said plurality of endothelial cells are further genetically transformed to express at least one marker polypeptide.

13. A transformed endothelial cell being genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

14. The transformed endothelial cell of claim 13, being further genetically transformed to express at least one marker polypeptide.

15. The transformed endothelial cell of claim 13, wherein said endothelial cell is derived from a source selected from the group consisting of a segment of a vein, and bone marrow progenitor cells, peripheral blood stem cells and circulating endothelial cells.

16. The transformed endothelial cell of claim 13, wherein said endothelial cell is derived from a source selected from the group

consisting of a human donor and an animal source.

17. The transformed endothelial cell of claim 13, wherein said cellular proliferating growth factor is selected from the group consisting of VEGF, acidic or basic FGF and HGF.

18. The transformed endothelial cell of claim 13, wherein said cellular adherence factor is DANCE or UP50.

19. A method of producing an artificial vascular graft, the method comprising the steps of:

- (a) genetically transforming endothelial cells to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor; and

- (b) culturing said endothelial cells within a synthetic tubular element having a luminal surface until sufficient endothelialization of said luminal surface is achieved.

20. The method of claim 19, wherein step (a) is effected by genetically transforming a first portion of said endothelial cells to express said at least one endothelial cell proliferating growth factor and genetically transforming a second portion of said endothelial cells to express said at least one cellular adherence factor.

21. The method of claim 19, wherein said step (a) precedes said step (b).

22. The method of claim 19, wherein said step (b) precedes said step (a).

23. The method of claim 19, wherein said endothelial cells are derived from a source selected from the group consisting of a segment of a vein, and bone marrow progenitor cells, peripheral blood stem cells and circulating endothelial cells.

24. The method of claim 19, further comprising the step of subjecting said luminal surface of said artificial graft to flow forces.

25. The method of claim 19, wherein said artificial graft is used in bypass surgery.

26. A nucleic acid expression construct comprising:

- (a) a first polynucleotide segment encoding an endothelial cell proliferating growth factor; and
- (b) a second polynucleotide segment encoding cellular adherence factor.

27. The nucleic acid expression construct of claim 26, further comprising at least one promoter sequence being for directing the expression of at least one of said first and said second polynucleotide segments.

28. The nucleic acid construct of claim 27, wherein said first polynucleotide segment is transcriptionally linked to said second

polynucleotide segment whereas said first and said second polynucleotide segment are under the transcriptional control of a single promoter sequence of said at least one promoter sequence.

29. The nucleic acid construct of claim 28, further comprising a linker sequence being interposed between said first and said second polynucleotide segments.

30. The nucleic acid construct of claim 29, where said linker sequence is selected from the group consisting of IRES and a protease cleavage recognition site.

31. The nucleic acid expression construct of claim 26, wherein said at least one promoter is functional in eukaryotic cells.

32. The nucleic acid expression construct of claim 26, wherein said at least one promoter is selected from the group consisting of a constitutive promoter, an inducible promoter and a tissue specific promoter.

33. The nucleic acid expression construct of claim 26, further comprising:

- (c) a first promoter sequence being for directing the expression of said first polynucleotide segment; and
- (d) a second promoter sequence being for directing the expression of said second polynucleotide segment.

34. The nucleic acid expression construct of claim 33, wherein said first promoter and said second promoter are selected from the group consisting of a constitutive promoter, an inducible promoter and a tissue specific promoter.

35. The nucleic acid expression construct of claim 34, wherein said inducible promoter are regulatable by same effector molecule.

36. The nucleic acid expression construct of claim 26, further comprising at least one additional polynucleotide segment encoding a marker polypeptide.

37. The nucleic acid expression construct of claim 36, wherein said marker polypeptide is selected from the group consisting of a

selection polypeptide and a reporter polypeptide.

38. The nucleic acid expression construct of claim 36, wherein said at least one additional polynucleotide segment is transcriptionally linked to said at least one of said first and said second polynucleotide segments.

39. The nucleic acid construct of claim 36, wherein said at least one additional polynucleotide segment is transcriptionally linked to said at least one of said first and said second polynucleotide segments via linker segment.

40. The nucleic acid construct of claim 39, wherein said linker sequence is selected from the group consisting of IRES and a protease

cleavage recognition site.

41. The nucleic acid construct of claim 36, wherein said at least one additional polynucleotide segment is translationally fused to at least one of said first and said second polynucleotide segments.

42. A nucleic acid expression construct system comprising:
- (a) a first nucleic acid expression construct including a first polynucleotide segment encoding an endothelial cell proliferating growth factor; and
 - (b) a second nucleic acid expression construct including a second polynucleotide segment encoding cellular adherence factor.

43. The nucleic acid expression construct system of claim 42, wherein at least one of said first and said second nucleic acid expression constructs further including at least one additional polynucleotide segment encoding a marker polypeptide.

44. The nucleic acid expression construct system of claim 42, further comprising at least one promoter sequence being for directing expression of at least one of said first and said second polynucleotide segment.

45. The nucleic acid expression construct system of claim 42, wherein said at least one promoter sequence is functional in eukaryotic cells.

46. The nucleic acid expression construct system of claim 42, wherein said at least one promoter sequence is selected from the group consisting of a constitutive promoter, an inducible promoter and a tissue specific promoter.

47. The nucleic acid expression constructs system of claim 43, wherein said marker polypeptide is selected from the group consisting of a selection polypeptide and a reporter polypeptide.

48. The nucleic acid expression constructs system of claim 43, wherein said at least one additional polynucleotide segment is transcriptionally linked to at least one of said first and said second polynucleotide segments.

49. A kit comprising a stand for engaging at least one tube, said at least one tube including a nucleic acid expression constructs system, said nucleic acid expression construct system including:

- (a) a first nucleic acid expression construct including a first polynucleotide encoding an endothelial cell proliferating growth factor; and
- (b) a second nucleic acid expression construct including a second polynucleotide encoding cellular adherence factor.

50. A kit comprising a stand for engaging at least one tube, said at least one tube including a nucleic acid expression construct, said nucleic acid expression construct including:

- (a) a first polynucleotide encoding an endothelial cell proliferating growth factor; and
- (b) a second polynucleotide encoding cellular adherence factor.

51. A method of producing genetically transformed endothelial cells, the method comprising the steps of:

- (a) obtaining endothelial cells from a source selected from the group consisting of a segment of a vein, and bone marrow progenitor cells; and
- (b) transforming said endothelial cells to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor concurrently.

52. A method of replacing or bypassing at least a portion of a vascular system of an individual, the method comprising the step of implanting into the vascular system of the individual an artificial vascular graft, so as to form a fluid communication between the vascular system and said artificial vascular graft, said artificial vascular graft including a synthetic tubular element having a luminal surface being coated with a plurality of endothelial cells being genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

53. The method of claim 52, wherein a first portion of said plurality of endothelial cells is genetically transformed to express said at least one endothelial cell proliferating growth factor and further wherein a second portion of said plurality of endothelial cells is genetically

transformed to express said at least one cellular adherence factor.

54. The artificial vascular graft of claim 52, wherein said synthetic tubular element is of an inner cross sectional area which is substantially equivalent to an inner cross sectional area of a blood vessel

55. The artificial vascular graft of claim 54, wherein said inner cross sectional area of said synthetic tubular element is within a range of about 7 to 700 mm².

ABSTRACT

The present invention provides an artificial vascular graft and methods of generating same. The artificial vascular graft includes a synthetic tubular element having a luminal surface being coated with a plurality of endothelial cells being genetically transformed to express at least one endothelial cell proliferating growth factor and at least one cellular adherence factor.

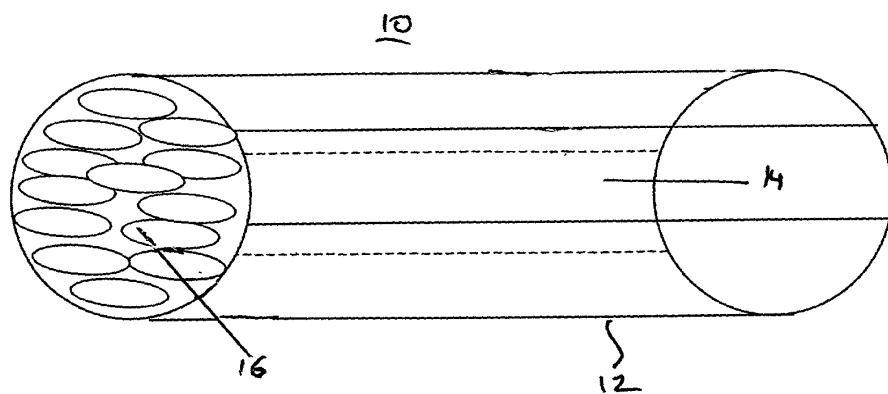
Fig 1

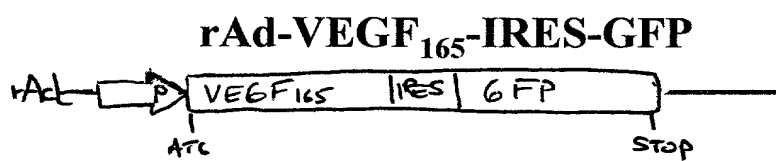
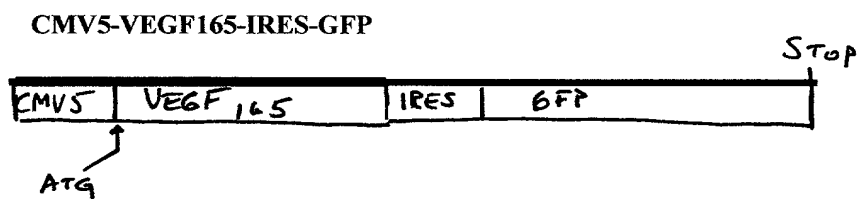
Fig 2aFig 2b

Fig 3a

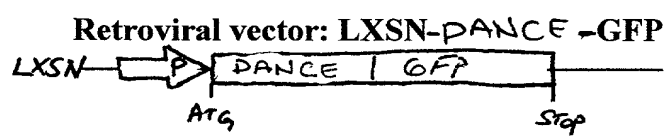


Fig 3b

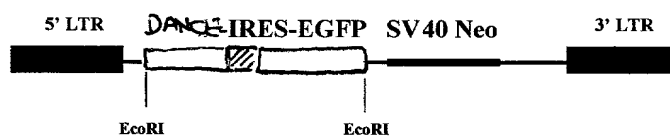
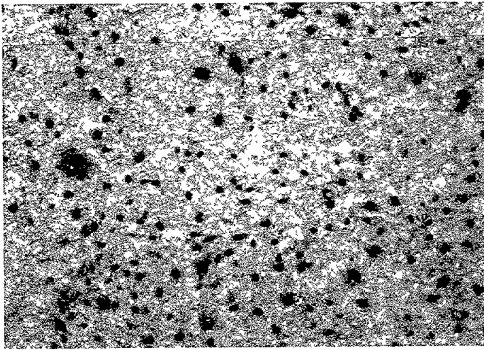
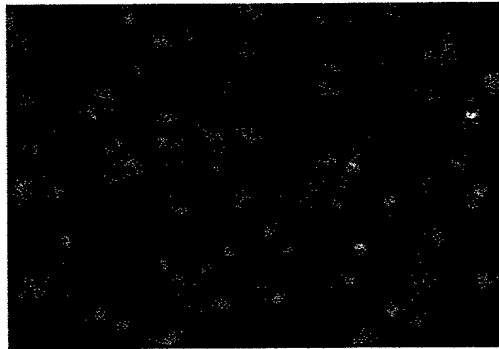
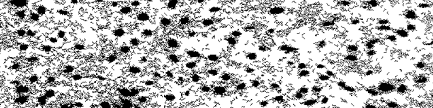
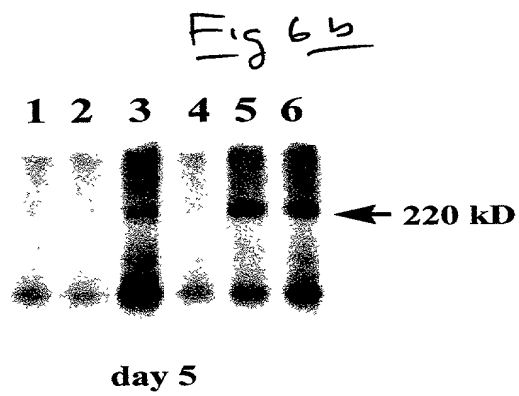
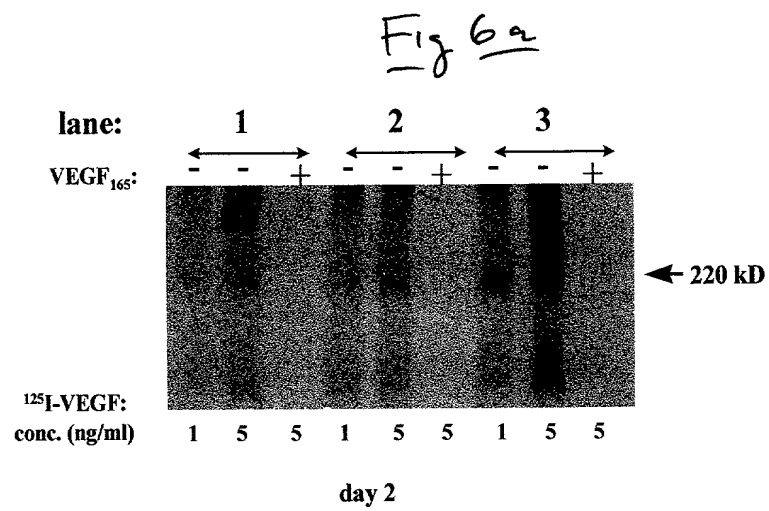


Fig 4aFig 4b





Combined Declaration For Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled ARTIFICIAL VASCULAR GRAFTS, AND METHODS OF PRODUCING AND USING SAME, the specification of which

(check one) ☒ is attached hereto.

☐ was filed on _____ as Application Serial No. _____ and was amended on _____. I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

NONE

(number) (Country) (Day, Month, Year Filed)

(number) (Country) (Day, Month, Year Filed)

(number) (Country) (Day, Month, Year Filed)

Priority Claimed

☐ ☐

Yes No

☐ ☐

Yes No

☐ ☐

Yes No

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States Application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

NONE

(Application Serial No.) (Filing Date)

Status

(patented, pending, abandoned)

(Application Serial No.) (Filing Date)

Status

(patented, pending, abandoned)

I hereby appoint the following attorneys, with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

Sol Sheinbein
Martin D. Moynihan

Registration No. 25,457
Registration No. 40,338

Address all Correspondence to:

G. E. EHRLICH (1995) LTD.
c/o ANTHONY CASTORINA
2001 JEFFERSON DAVIS HIGHWAY
SUITE 207
ARLINGTON, VIRGINIA 22202


Direct all telephone calls & faxes to:

Phone (703) 415-1581
Fax (703) 415-4864

Attorney Docket: 20002
page 2 of 2

Continuation of Combined Declaration For Patent Application and Power of Attorney

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statement may jeopardize the validity of the application of any patent issued thereon.

*FULL NAME OF SOLE OR FIRST INVENTOR MOSHE FLUGELMAN	INVENTOR'S SIGNATURE 	DATE 16-July-2000
RESIDENCE 80 Hanasi Ave. Haifa, ISRAEL	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS 80 Hanasi Ave. Haifa, ISRAEL		

*FULL NAME OF SECOND INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

*FULL NAME OF THIRD INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

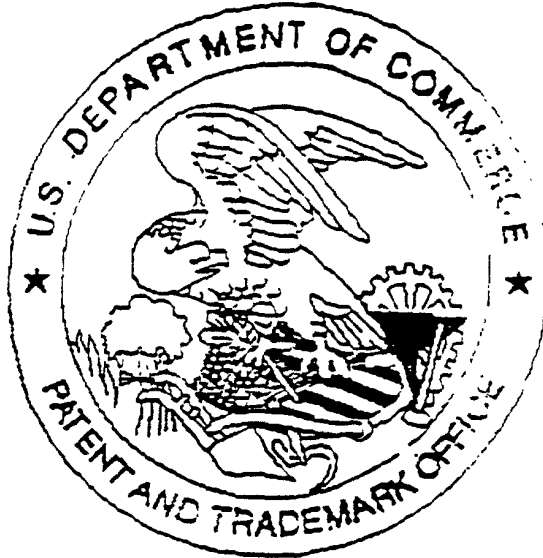
*FULL NAME OF FOURTH INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

*FULL NAME OF FIFTH INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

*FULL NAME OF SIXTH INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

*FULL NAME OF SEVENTH INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE	CITIZENSHIP ISRAELI	
POST OFFICE ADDRESS		

United States Patent & Trademark Office
Office of Initial Patent Examination -- Scanning Division



Application deficiencies were found during scanning:

☐ Page(s) _____ of _____ were not present:
for scanning. (Document title)

☐ Page(s) _____ of _____ were not present
for scanning. (Document title)

☒ Scanned copy is best available. small entity, drawings,
declaration